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DNA Cloning

- DNA cloning is a technique that allows for:
 - Amplifying a DNA segment into many, many copies in a biological system
 - **Expressing** a gene inside a biological system
- Recombinant DNA is composed of the *gene of interest* (encoding a protein or non-coding RNA) and a *vector* formed using restriction endonucleases and ligase
 - > Cut, Join, Insert into a cell

Cloning means that you make several copies of one thing

- A clone is a genetically identical population (organisms, cells, viruses, DNA)
- Every member of the population is derived from a single cell, virus, or DNA molecule
- Restriction endonuclease cut in specific fragments of 4-8 bp palindromic sequences
- There are 2 types of cut can be produced:
 - Staggered (off-center): enzymes cut the two DNA strands at different positions generating sticky or cohesive ends where the fragments have short single stranded overhangs at each end
 - ✓ Can form H-bonds with *complementary* sequences, so can be used in cloning
 - *Blunt:* enzymes cut at the *same position* on both strands giving blunt-ended fragments
- *Vector:* It is the *carrier* where the gene of interest is added
 - Bacterial plasmids are considered excellent vectors for cloning (cloning vectors) or expression (expression vectors) where it is a small circular DNA separated from the main bacterial chromosome
- Features of plasmid cloning vectors:
 - Their own origin of replication (OriC) that allows them to replicate independently of the bacterial chromosome
 - > A selectable gene such as an <u>antibiotic resistance gene for ampicillin</u> antibiotic
 - > A *restriction site* that allows for insertion of the DNA segment of interest
- The gene of interest is added and joined with the vector by DNA ligase
 - DNA ligase: It covalently joins DNA ends by catalyzing the ATP-dependent formation of permanent phosphodiester bonds between 3' hydroxyl of one strand and 5' phosphate of the other strand
- The vector must be previously cut by the <u>same restriction endonuclease</u> that the gene of interest cut by
 - > To make sticky ends complementary to that in the gene of interest
- DNA libraries are 2 types:
 - > Genomic library: It contains all the elements of the genome (exons, introns, telomere, ...)
 - ✓ A genome is *cleaved* by the <u>same restriction endonuclease</u> as that used for the vector and each fragment is *ligated* into a vector, then each plasmid is *transferred int a bacterial cell* (each cell has 1 plasmid) and can grow into millions of cells and each cell can *make multiple copies* of every plasmid ending up with billions of copies of plasmid with a specific DNA fragment.







> cDNA Library

- *mRNAs* are isolated and *reverse transcribed* by reverse transcriptase into a cDNA molecule that is *replicated* by DNA polymerase to form a double-stranded cDNA, then *synthetic linkers* containing a restriction site are ligated to the ends of the cDNAs and digested with the *restriction endonuclease* to form overhangs and then cDNAs are *cloned* into a plasmid
- ✓ Contains only the elements present in the *mature mRNA* (exons)
- ✓ *Only transcribed* genes can appear

Analysis of transcriptional regulatory sequences

- It is the study of the effect of different conditions on the regulation and level of expression
- It requires a *reporter gene* such as *luciferase gene*
 - Luciferase is an enzyme in fruit flies that convert luciferin into oxyluciferin which is glowing
- The reporter gene must be inserted to the plasmid containing the gene of interest, where *only the regulatory sequences* (promoter, PPE, enhancers and silencers) are *upstream* to it
 - The plasmid is transfected (inserted) into cells, and the expression level of luciferase (instead of the original gene itself) is *measured via the strength of the <u>light signal</u>*





Analysis of gene expression

• It is done by measuring RNA levels by basic (northern blotting, in situ hybridization), advanced (real-time PCR, DNA microarray) and very advanced methods (RNA-seq)

Northern blotting

- This is done exactly like Southern blotting with some differences:
 - **RNA** from cells is isolated instead of DNA and fractionated based on size by gel electrophoresis
 - RNA molecules are transferred onto a membrane and targeted by a *labeled DNA probe* with a sequence that is complementary to a specific RNA molecule

Western blotting: For detection of *proteins* via SDS-PAGE, and using *antibodies* instead of probes

- It can tell us if the gene is *expressed or not*
- If expressed it can tell us the *size* (via the *position*) and *amount of RNA* (via *band intensity*)
 - If you studied one gene only and 2 bands (with different sized) appeared in the same samples that could be variety in transcription due to *different promoters*, or different RNA processing (*alternative splicing* or *polyadenylation*)



- > In this case, house-keeping genes must be assessed also to be sure that the sample is taken properly
- House-keeping genes are genes expressed in all cells, in the same levels (constant expression, do not change) such as histones, actin, tubulin and some metabolic enzymes



In situ hybridization

- In situ hybridization methods reveals the *distribution* of specific RNA molecules in cells in *tissues*
- RNA molecules can hybridize when the tissue is incubated with a complementary *DNA or RNA probe*
- In this way the patterns of differential gene expression can be observed in tissues, and the location of specific RNAs can be determined in cells
- We use in situ hybridization to know in which cells the gene is expressed, and immunohistochemistry to know where the protein is concentrated



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Reverse transcriptase Real time qPCR

- Quantitative reverse transcriptase RT-qPCR of mRNA is another way of relative quantitation of RNA expression is by converting RNA into cDNA followed by PCR in the presence of SYBR green
 - The *higher the amount* of RNA = higher amount of cDNA, the sooner detected
- To compare expression level between 2 tissues, *house keeping* genes analysis is required to ensure proper sample taking







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SPAD count



- Detection of **SARS-Co-2** is done by qPCR
 - We must analyze the expression of at least 1 viral gene and 1 human gene
 - The analysis of *human gene expression* is to ensure proper sample taking



- The study of -omics
 - Genomics: The study of genomes and DNA sequence
 - > Transcriptomics: They study of RNA produced by transcription, mainly by DNA microarray
 - Proteomics: The study of proteins \geq
 - Metabolomics: The study of metabolites produced from enzymes \geq

DNA microarrays

- DNA microarrays are solid surfaces (glass microscope slides or chips) spotted with up to tens of thousands of DNA fragments in an area the size of a fingernail. Where *the* exact sequence and position of every DNA fragment on the array is known
- mRNAs are extracted from cells (cancer and normal for example) and converted to cDNAs, which are labeled with a *fluorescent dye*, then cDNAs hybridize to a DNA corresponding to distinct human genes
 - > The relative level of expression of each gene is indicated by the **intensity of** fluorescence at each position on the microarray
 - > Can be used to compare the types of genes expressed between different cell types



RNA sequencing (RNA-seq)

- Cellular RNA is reverse transcribed to *cDNAs*, which are subjected to next-generation sequencing
 - The relative amount of initial mRNA is indicated by the <u>frequency</u> at which its sequence of cDNA is read



- *RNA-seq* can be used to characterize *novel transcripts*, identify splicing *variants* and profile the *expression levels* of all transcripts
 - > *Microarrays* are limited to detect transcripts corresponding to *known genomic sequences*





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